Effect of Acrylamide on the Tongue Musculature of Adult Male Albino Rat and the Possible Protective Role of Vitamin E: Light and Electron Microscopic Study

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Abstract

Background: Acrylamide is a potent toxin and carcinogen, its hazards result from its increased accumulation in the process of cooking food materials.

Aim of Study: This study aimed to demonstrate the effects of acrylamide on the skeletal muscle fibers of adult male albino rat tongue for two different durations, using light and electron microscopic study and to investigate the possible protective effect of Vitamin E co-administration.

Material and Methods: Sixty adult male albino rats were divided into six groups, each group included ten rats. Group I (control). Group II (Sham control). Group III received acrylamide for 20 days. Group VI received acrylamide for 40 days. Group V received acrylamide and Vitamin E, with 30 minutes interval between the two doses, for 20 days. Group VI received acrylamide and Vitamin E with 30 minutes interval between the two doses, for 40 days. All medications were given orally by gastric gavage. At the end of the experiment the tongue was dissected out for light and electron microscopic studies.

Results: Light microscopic examination revealed that acrylamide lead to skeletal muscle damage in rat tongue. In the form of, fragmentations, splitting and inflammatory cellular infiltrations. Electron microscopic study of Group III revealed degeneration of mitochondria, these changes became worse with prolonged duration of exposure. This toxic effect of acrylamide could be minimized when Vitamin E is given concomitantly with it.

Conclusion: Acrylamide lead to histological changes in rat tongue. Vitamin E minimized this changes due to its antioxidant effect.

Key Words: Acrylamide – Tongue – Histology – Electron microscope Vitamin E.

Introduction

ACRYLAMIDE, one of the major environmental public health problems, found in carbohydrate-rich food prepared at high temperatures which are widely consumed by humans. Consumption of these foods may result in significant human exposure to acrylamide [1]. Acrylamide is a low molecular weight, water soluble vinyl monomer; these properties facilitate its rapid absorption and distribution through body [2].

Acrylamide has been extensively investigated and has a large database of very complex toxicity. The results of the studies indicate that acrylamide is carcinogenic in rodents and produces toxic effects on the different body organs, e.g. kidney, spleen and liver [3,4].

El-Sayyad et al., [5] reported that sections of the skeletal muscles of tongue of rats treated with acrylamide showed vacuolated spaces and many of spotty necrotic zones. Partial hyaline necrosis was detected in the majorities of the skeletal muscle fibers. Focal necrotic zones with dispersed nuclei were detected. Loosely separated skeletal muscle fibers associated with lacking of their regular arrangement and densely collected inflammatory cells in between were observed.

The administration of acrylamide was associated with a dose related increase in squamous cell papilloma of the tongue and combined squamous cell papilloma and carcinoma of the tongue in rats [6].

Exposure to acrylamide resulted in reduction of lingual keratinization, atrophy of different tongue papillae and degeneration of the apical taste buds of fungiform papillae and abundant leucocytic infiltrations appeared in the lamina propria of the tongue of experimental rats [7].
Longitudinal sections of the skeletal muscles of the tongue of acrylamide treated rats showed disruption of Z line, partial loss of myofilaments, marked aggregation of mitochondria and vacuolated mitochondria also appeared [8].

Oxidative stress and apoptosis play an important role in acrylamide-induced damage in skeletal muscle fibers of the tongue. Prasad, [9] reported that acrylamide-induced cytotoxicity was relevant to oxidative stress, the cytotoxic properties of acrylamide lead to generation of Reactive Oxygen Species (ROS) causing cytotoxic and genotoxic effects.

Some strategies have been postulated to reduce acrylamide mediated cytotoxicity by using natural antioxidant like Vitamin E. Vitamin E can protect cellular structures against damage from free radicals such as peroxy radical, hydroxyl radical and superoxide. It has protective effect from oxidation products such as Malondialdehyde (MDA) and hydroxynonenal. Vitamin E was utilized to prevent the toxic effects of acrylamide on the testes [10].

The aim of this study is to investigate the effect of acrylamide on the skeletal muscle fibers of tongue of adult male albino rat and the possible protective role of Vitamin E co-administration, for a short duration (twenty days) and a long duration (forty days) by using light and electron microscopic study.

Material and Methods

Material:

Animals:

The research was done in 2017. This study was performed using 60 adult male albino rats weighing 180-220gm. The rats were obtained from Animal House, Faculty of Medicine, Cairo University. The rats were acclimatized in the laboratory for a period of two weeks before carrying out the experiment. They were housed in cages, five rats/cage under standard laboratory and environmental conditions. They were allowed free access to food and distilled water. All the animals were cared for in accordance with the institutional ethical guidelines and protocol was approved by the Kasr Al-Einy Experimental Ethics committee. The rats were randomly divided into six experimental groups; each group consists of ten rats:

- **Group I: (Normal control):** Rats received no medications.
- **Group II: (Sham control):** Rats received normal saline solution and Vitamin E dissolved in corn oil orally once daily by gavage, with 30 minutes interval between the two doses, for 40 days.
- **Group III:** Rats received acrylamide orally once daily by gavage for 20 days [11].
- **Group IV:** Rats received acrylamide orally once daily by gavage for 40 days [8].
- **Group V:** Rats received acrylamide and Vitamin E orally once daily by gavage, with 30 minutes interval between the two doses, for 20 days [11].
- **Group VI:** Rats received acrylamide and Vitamin E orally once daily by gavage, with 30 minutes interval between the two doses, for 40 days [8].

Chemicals:

Acrylamide (99.9 purity), obtained from sigma chemical Co. Each 20mg of acrylamide was dissolved in 5ml saline solution and given orally by gastric gavage in a daily dose of 20mg/kg body weight. Each rat was given the calculated dose of Vitamin E, (Capsules 400mg/capsule) obtained from sigma chemical Co. Each Vitamin E capsule 400mg was dissolved in 20ml of corn oil and given orally by gastric gavage in a daily dose of 100mg/kg body weight [12]. Each rat was given the calculated dose.

Rats from Groups (I and II) were divided into two subgroups (5 rats in each subgroup), 5 rats were sacrificed after 20 days and 5 rats were sacrificed after 40 days, rats from Groups (III and V) were sacrificed after 20 days, rats from Groups (IV and VI) were sacrificed after 40 days.

At the end of the experiment, the tongue was dissected, all samples were collected for light and electron microscopic studies.

Methods:

Light microscopic study:

For histological examination, coronal sections of the tongue will be stained with hematoxylin and eosin (H & E), Masson’s trichrome and P.A.S. stains.

Histomorphometric study:

- Percentage of collagen fibers in Masson’s trichrome stained sections.
- PAS optical density in PAS stained sections.

Electron microscopic study:

The ultrastructural study of tongue samples will be carried out with transmission electron microscope.
Statistical analysis:

Data was expressed as mean value ±SD. Comparisons between two related groups were carried out by paired t-test while comparisons between more than two groups were carried out by ANOVA test (analysis of variance) followed by Tukey’s test.

Statistical analysis was done for:
A- The percentage of collagen fibers in all groups.
B- The mean PAS optical density in all groups.

Results

Light microscopic results:

Groups I & II (normal and sham control groups):

The light microscopic results of the normal and sham control groups were similar.

The histological sections of the rat tongue of the normal and sham control groups stained with haematoxylin and eosin showed bundles of skeletal muscle fibers running in different directions, the skeletal muscle fibers running longitudinally arranged in bundles or fascicles of muscle fibers separated by perimysial connective tissue, the muscle fibers appeared parallel, long, cylindrical and non-branching with acidophilic crossly striated sarcoplasm. The nuclei were multiple, elongated and peripheral in position under the sarcolemma. The fibers were connected together by connective tissue endomysium Fig. (1A).

The skeletal muscle fibers running transversely appeared polygonal with acidophilic myofibrils and peripherally located nuclei. The endomysial connective tissue among the muscle fibers showed some blood vessels Fig. (1A).

Sections stained with Masson's trichrome visualized a normal pattern of collagen deposition around skeletal muscle fibers Fig. (1B).

Sections stained with PAS, the skeletal muscle fibers were shown to have a strong positive PAS reaction in the form of small red granules filling their sarcoplasm denoting considerable amounts of glycogen Fig. (1C).

Group III (animals receiving acrylamide for short duration):

In the histological sections of the rat tongue of Group III (after 20 days of acrylamide administration), the skeletal muscle fibers showed abnormal wavy course of the muscle fibers with splitting of the myofibers and fragmentation of the sarcoplasm Figs. (2A-C, 3A). In addition, dilated congested blood vessels with fibrosed walls Fig. (2B) and mononuclear cellular infiltration appeared in the connective tissue perimysium Fig. (4) and darkly stained pyknotic nuclei were also detected Fig. (3A). Using Masson's trichrome stain technique the deposition of collagen fibers increased around the skeletal muscle fibers Fig. (3A). In specimens stained with PAS, the skeletal muscle fibers showed faint PAS reaction indicating reduced glycogen content Fig. (3B).

Group IV (animals receiving acrylamide for long duration):

In the histological sections of the rat tongue of Group IV (after 40 days of acrylamide administration), the splitting and fragmentation of the myofibers were markedly increased. Dense pyknotic nuclei and remnants of nuclei were also detected Fig. (4A). Markedly dilated and congested blood vessels with fibrosed walls were observed Fig. (4A). Using Masson's trichrome stain technique the deposition of collagen fibers was markedly increased around the skeletal muscle fibers Fig. (4B). In specimens stained with PAS, the skeletal muscle fibers showed markedly faint PAS reaction indicating markedly reduced glycogen content Fig.

Group V (animals receiving acrylamide and Vitamin E for short duration):

The histological sections of the rat tongue of Group V (after 20 days of acrylamide and Vitamin E administration), showed mild focal histological changes. Few fibers still have wavy course with mild splitting and fragmentation Fig. (5A). In Masson's trichrome stained sections, there was mild increase in the collagen fibers deposition around the skeletal muscle fibers Fig. (5B). Sections stained with PAS revealed a moderate glycogen content as demonstrated by their moderately positive PAS reaction Fig. (5C).

Group VI (animals receiving acrylamide and Vitamin E for long duration):

The histological sections of the rat tongue of Group VI (after 40 days of acrylamide and Vitamin E administration), showed moderate focal histological changes. Some fibers still have wavy course with moderate splitting and fragmentation Fig. (6A). Using Masson's trichrome stain technique the deposition of collagen fibers was moderately increased around the skeletal muscle fibers Fig. (6B). In specimens stained with PAS, the skeletal muscle fibers showed moderately faint PAS reaction indicating reduced glycogen content Fig. (6C).
Electron microscopic results:

Groups I & II (normal and sham control groups): The electron microscopic results of the normal and sham control groups were similar. Using the electron microscope, the sarcoplasm appeared filled with myofibrils arranged parallel to the long axis of the myofiber. The myofibrils showed regular arrangement of alternating light (I) and dark (A) bands. Z line was seen bisecting the light band. Sarcomeres were seen between two successive Z lines Fig. (7A).

Group III (animals receiving acrylamide for short duration): Using the electron microscope, micrographs of skeletal muscle fibers of Group III showed aggregation of mitochondria of different size and shape in the subsarcolemmal and intermyofibrillar spaces. Degeneration of parts of myofibrils and disruption of Z lines were observed. Fusion of some mitochondria to form giant and vacuolated mitochondria appeared Fig. (7B).

Group IV (animals receiving acrylamide for long duration): Using the electron microscope, micrographs of skeletal muscle fibers of Group IV, areas of marked myofibrillar degeneration with loss of sarcomere pattern and disruption of Z lines were observed. Marked aggregations of giant and vacuolated mitochondria in the subsarcolemmal and intermyofibrillar spaces were detected Fig. (7C).

Group V (animals receiving acrylamide and Vitamin E for short duration): Using the electron microscope, micrographs of skeletal muscle fibers of Group V showed mild degeneration of myofibrils, nearly normal patterns of sarcomeres with parallel arrangement, light and dark bands and Z lines were preserved Fig. (8A).

Group VI (animals receiving acrylamide and Vitamin E for long duration): Using the electron microscope, micrographs of skeletal muscle fibers of Group VI showed areas of moderate myofibrillar degeneration and disruption of Z lines. Moderate aggregations of giant and vacuolated mitochondria in the subsarcolemmal and intermyofibrillar spaces were detected Fig. (8B).
Fig. (2): A- A photomicrograph of a section of tongue of a rat from group III showing disrupted wavy course of the skeletal muscle fibers with splitting of some fibers (Sp), fragmentation (Fr) of the sarcoplasm, dilatation (Dv) of blood vessels and cellular infiltration (blue arrows). (Hx & E; X100) B- A photomicrograph of a section of tongue of a rat from group III, showing disrupted wavy course of the skeletal muscle fibers with splitting of some fibers (Sp), fragmentation (Fr) of the sarcoplasm and congestion of blood vessels (Cv). (Hx & E; X200). C- A photomicrograph of a section of tongue of a rat from group III, showing disrupted wavy course of the skeletal muscle fibers with splitting of some fibers (Sp), fragmentation (Fr) of the sarcoplasm. Notice the presence of darkly stained pyknotic nuclei (P). (Hx & E; X400).

Fig. (3): A- A photomicrograph of a section of tongue of a rat from Group III, showing increased collagen fibers deposition (white arrow) between the skeletal muscle fibers running in different directions and around blood vessels. (Masson's trichrome; X400). B- A photomicrograph of a section of tongue of a rat from Group III exhibiting a faint PAS reaction of the skeletal muscle fibers (green arrow). (PAS; X400).
Fig. (4): A- A photomicrograph of a section in the tongue of a rat from Group IV, showing marked disrupted wavy course of the skeletal muscle fibers with splitting of some fibers (Sp), marked fragmentation (Fr) of the sarcoplasm, markedly dilated and congested blood vessels (Cv). Notice the presence of darkly stained pyknotic nuclei (P). (Hx & E; X400). B- A photomicrograph of a section of tongue of a rat from Group IV, showing marked increase in collagen fibers deposition (white arrow) around the skeletal muscle fibers running in different directions. (Masson's trichrome; X400). C- A photomicrograph of a section in the tongue of a rat from Group IV exhibiting a markedly faint PAS reaction of the skeletal muscle fibers (green arrow). (PAS; X400).

Fig. (5): A- A photomicrograph of a section of tongue of a rat from Group V, showing nearly normal skeletal muscle fibers running longitudinally. Minute fragmentations (Fr) occasionally found. (Hx & E; X400). B- A photomicrograph of a section of tongue of a rat from the Group V, showing nearly normal pattern of collagen fibers deposition (white arrow) around the skeletal muscle fibers running in different directions. (Masson's trichrome; X400). C- A photomicrograph of a section of tongue of a rat from the group V exhibiting a moderately positive PAS reaction of the skeletal muscle fibers. (PAS; X400).
Fig. (6): A- A photomicrograph of a section of tongue of a rat from group VI, showing moderate skeletal muscle fibers splitting (Sp) and moderate fragmentations (Fr). Fatty infiltration (Fi), dilatation (Dv) and darkly stained pyknotic nuclei (P) also appeared. (Hx & E; X400). B- A photomicrograph of a section of tongue of a rat from the Group VI, showing nearly normal pattern of collagen fibers deposition (white arrow) between the skeletal muscle fibers running in different directions. (Masson's trichrome; X400). C- A photomicrograph of a section of tongue of a rat from the Group VI exhibiting a moderately faint PAS reaction of the skeletal muscle fibers. (PAS; X400).

Fig. (7): A- An electron photomicrograph of a section of tongue of a rat from normal control group showing parallel arrangement of myofibrils with light bands (Lb) and dark bands (Db). Sarcomeres (S) are seen between two successive Z lines (Z). (X10000). B- An electron photomicrograph of a section of tongue of a rat in Group III, showing aggregation of giant mitochondria (Mg) and vacuolated mitochondria (Mv), degeneration of myofibrils (Dg), notice the disruption of Z lines and the loss of sarcomere pattern. (X10000). C- An electron photomicrograph of a section of tongue of a rat in Group IV, showing marked degeneration of myofibrils (Dg), marked aggregations of vacuolated mitochondria (Mv). Notice the loss of sarcomere pattern. (X10000).
Histomorphometric and statistical results:

A- Statistical analysis of the mean area percent of collagen fibers (Table 1) and Histogram (1):

Statistical analysis of data showed that there was no statistical significant difference in the mean percent area of collagen fibers between the normal Group I and sham control Group II. The mean area percent of collagen fibers in Group III was highly significant different compared to the control group and in Group IV the difference was highly significant increased when compared to the control group. In Group V, the mean area percent of collagen fibers was non significant compared to control group but was highly significant different compared with Group III and Group IV. In Group VI the difference was non significant compared to control group but was highly significant different compared with Group IV. The F-ratio high numerical value (11.986) indicates a high overall significance among the different animal groups.

B- Statistical analysis of the mean PAS optical density (Table 2) and Histogram (2):

Statistical evaluation of the mean values of the PAS optical density in the different experimental groups showed no statistically significant difference between the normal control Group I and sham control Group II. On the other hand, the mean glycogen content in Group III showed highly significant different compared to the control group and in Group IV the difference was highly significant decreased when compared to the control group. In Group V, the mean glycogen content was non significant compared to control group but was significant different compared with Group III and was highly significant different compared with Group IV. In Group VI the difference was non significant compared to control group and was significant different compared with Group IV. The F-ratio high numerical value (12.426) indicates a high overall significance among the different animal groups.
Table (1): The mean area percent of collagen fibers in all groups using ANOVA test.

<table>
<thead>
<tr>
<th>Groups</th>
<th>Mean ± SD</th>
<th>p-value</th>
<th>F-ratio</th>
</tr>
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<tbody>
<tr>
<td>Group I (Control group)</td>
<td>2.38±0.41</td>
<td>–</td>
<td>–</td>
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<tr>
<td>Group II (Sham control group)</td>
<td>2.42±0.24</td>
<td>0.982*</td>
<td>0.007***</td>
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<tr>
<td></td>
<td></td>
<td>&lt;0.0001****</td>
<td>0.475*****</td>
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<tr>
<td>Group III (Acrylamide short duration)</td>
<td>7.51±1.60</td>
<td>0.006*</td>
<td>0.007**</td>
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<tr>
<td></td>
<td></td>
<td>0.032****</td>
<td>0.141*****</td>
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<tr>
<td>Group VI (Acrylamide long duration)</td>
<td>12.99±5.42</td>
<td>&lt;0.0001*</td>
<td>&lt;0.0001**</td>
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<tr>
<td></td>
<td></td>
<td>0.004***</td>
<td>&lt;0.0001****</td>
</tr>
<tr>
<td>Group V (Acrylamide &amp; Vit. E short duration)</td>
<td>3.63±0.77</td>
<td>0.461*</td>
<td>0.475**</td>
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<td></td>
<td></td>
<td>&lt;0.0001****</td>
<td>0.440****</td>
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<tr>
<td>Group VI (Acrylamide &amp; Vit. E long duration)</td>
<td>4.95±0.69</td>
<td>0.140*</td>
<td>0.146**</td>
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<tr>
<td></td>
<td></td>
<td>&lt;0.0001****</td>
<td>0.440****</td>
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p-value is significant ≤0.05. ** : With group 1. *** : With group 4. **** : With group 5. #SD: Standard deviation.
The F-ratio: Overall significance among the different animal groups.

Table (2): The mean PAS optical density in all groups using ANOVA test.

<table>
<thead>
<tr>
<th>Groups</th>
<th>Mean ± SD</th>
<th>p-value</th>
<th>F-ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group I (Control group)</td>
<td>13.66±4.77</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>Group II (Sham control group)</td>
<td>13.57±4.78</td>
<td>0.964*</td>
<td>0.002***</td>
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<tr>
<td></td>
<td></td>
<td>&lt;0.0001****</td>
<td>0.162*****</td>
</tr>
<tr>
<td>Group III (Acrylamide short duration)</td>
<td>6.42±0.54</td>
<td>0.002*</td>
<td>0.002**</td>
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<tr>
<td></td>
<td></td>
<td>0.049*****</td>
<td>0.281********</td>
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<tr>
<td>Group VI (Acrylamide long duration)</td>
<td>1.83±0.61</td>
<td>&lt;0.0001*</td>
<td>&lt;0.0001**</td>
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<td></td>
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<td>0.034***</td>
<td>0.0001****</td>
</tr>
<tr>
<td>Group V (Acrylamide &amp; Vit. E short duration)</td>
<td>10.65±0.91</td>
<td>0.150*</td>
<td>0.162**</td>
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<tr>
<td></td>
<td></td>
<td>&lt;0.0001****</td>
<td>0.066******</td>
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<tr>
<td>Group VI (Acrylamide &amp; Vit. E long duration)</td>
<td>7.98±0.93</td>
<td>0.082*</td>
<td>0.086**</td>
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<tr>
<td></td>
<td></td>
<td>0.046****</td>
<td>0.066******</td>
</tr>
</tbody>
</table>

p-value is significant ≤0.05. ** : With group 1. *** : With group 4. **** : With group 5. #SD: Standard deviation.
The F-ratio: Overall significance among the different animal groups.

**Discussion**

In the present study, light microscopic examination of Group III, in which rats received acrylamide for 20 days, showed histopathological changes as abnormal wavy course of the skeletal muscle fibers with splitting of the myofibers and fragmentation of the sarcoplasm. These findings are in agreement with the results of Cullen et al., (2001) [13] that explained the splitting of muscle fibers as an adaptive response, which occurs when the fiber reaches a critical size at which supply of oxygen and exchange of metabolites are no longer efficient. In the current study, vascular congestion and perimuscular and perivascular fibrosis were detected that coincides with the results of our study. Also, Maronpot et al., (2015) [15] noticed in their study muscle fiber degeneration accompanied by inflammatory cell responses including mononuclear cellular infiltration and fibrosis.

Stevens and Lowe (2000) [16] explained that mononuclear cellular infiltration and fibrosis could be due to the release of certain mediators during degeneration of myocytes which initiate inflammatory reaction and attract inflammatory cells. This explanation was confirmed also by Nie and Hou (2012) and agreed with the findings of our study.

In our study, dense pyknotic nuclei and remnants of nuclei were observed. This agrees with the results of Prasad (2012) [9], which stated that exposure to acrylamide induces cytotoxic and genotoxic effects and promotes apoptosis. Weiss et al., (2013) [17] also noticed these changes and...
mentioned that they were associated with increased oxidative stress.

In Group IV, the previous histopathological changes were exaggerated with prolongation of the duration of exposure, as regard the fragmentation and splitting of the skeletal muscle fibers, in between the skeletal muscle fibers, the mononuclear cellular infiltration and vascular congestion, dilatation and fibrosis. This was confirmed previously by Shinomol et al., (2013) [13] and Rashay and Fatma (2015) [8], as they stated that the toxic effects of acrylamide is a dose dependent which increased by increasing the dose or the duration.

In our study, electron microscopic examination of skeletal muscle fibers of Group III showed degenerated parts of myofibrils, these results agrees with El-Sayyad et al., (2011) [5] and Shipp et al., (2006) [18], that reported that sections of the skeletal muscles of tongue of rats treated with acrylamide showed focal necrotic zones associated with lacking of their regular arrangement. Fusion of some mitochondria to form giant ones and vacuolation of others, disruption of Z line and partial loss of myofilaments were observed, these results agrees with the findings of Ghanayem et al., (2005) [19] and Lakshmi et al., (2012) [20]. Lakshmi et al., (2012) [20] stated that mitochondrial dysfunction has an important role in acrylamide induced toxicity.

By 40 days of exposure in Group IV, these changes were markedly exaggerated. These results and effect of acrylamide on mitochondria was described previously by El-Sayyad et al., (2011) [5] and Yang et al., (2005 [21]). It also stated that morphological mitochondrial abnormalities are most frequently associated with defects of the respiratory chain. Subsarcolemmal accumulations of mitochondria and their proliferation were also found and explained by Youssef (2006) [22] as an attempt by the cell to compensate for the respiratory chain defect.

The findings of the current study are mostly related to the oxidative stress effect of acrylamide on the skeletal muscle and it coincides with the study of Hori et al., (2013) [23] who stated that destruction of muscle fibers with oedema and inflammatory cell infiltration are prominent features of oxidative stress effect on skeletal muscle fibers. Their findings were also observed in our study as there were degenerated segments of the muscle fibers and mononuclear cellular infiltration in Group III and increased more in Group IV.

In our results, apoptosis which is presented by appearance of dense pyknotic nuclei and remnants of nuclei could be explained also by the oxidative stress damage effect of acrylamide on skeletal muscle which proved by Sumizawa et al., (2009) [24] and Lakshmi et al., (2012 [20]). They stated that oxidative stress and apoptosis play an important role in acrylamide induced toxicity.

In the current study, there are statistically significant increase in collagen fibers deposition and decrease in PAS optical density in Group III and IV than the control group and Group IV than Group III. These results coincide with Youssef (2006) [22], Catalgol et al., (2009) [25] and Huda et al., (2017) [26] who explained that these findings are indicators of oxidative stress in skeletal muscle fibers of acrylamide exposed rats that increased by prolongation of the duration of exposure.

In the present study, Vitamin E was administrated in combination with acrylamide for two different durations. The histological picture was improved in Group V (treated group for 20 days) and Group VI (treated group for 40 days) than the non-treated Groups (III and IV respectively). Histological sections of the skeletal muscle fibers of Vitamin E treated rats of Group V and VI revealed an apparently preserved architecture. This finding comes in accordance with Rasha and Fatma (2015) [8]. Also Nisreen et al., (2017 [27]) suggested that Vitamin E can eliminate free radical damage that induces apoptosis. Bermudz et al., (2007) [28] added that the antiapoptotic effects of Vitamin E are primarily associated with its antioxidant activity. Vitamin E has also been shown to induce protective effects and prevent apoptosis in many experimental model systems.

In the present study, Vitamin E treated Group V appeared with minimal amount of collagen fibers deposition in Masson trichrome stained sections and a positive reaction was observed in PAS stained sections, these coincides with Hoda et al., (2017), who stated that Vitamin E supplementation with acrylamide significantly protected and prevented oxidative stress induced degenerative changes and promotes regeneration.

In our study, the electron microscopic examination of the skeletal muscle fibers of Group V and Group VI showed nearly preserved normal structure. Most of cell organelles were intact. Mild focal degeneration occasionally found. These findings were confirmed by Rasha and Fatma (2015 [8]).
In the present work, there are statistically significant decrease in collagen fibers deposition and increase in PAS optical density in Group V and VI than Group III than Group IV respectively. These findings are indicator of oxidative stress in skeletal muscle fibers of acrylamide exposed rats that decreased by Vitamin E co-administration. These results coincide with the findings of Shinomol et al., (2013) [13] and Rasha and Fatma (2015) [8].

From the present study, it could be concluded that, acrylamide induced skeletal muscle damage in rat tongue which increasing by prolonged duration of exposure. Acrylamide induced oxidative stress is the implicated mechanism of such historical changes. This toxic effect of acrylamide could be decreased by administration of Vitamin E given concomitant with it. This protective role of Vitamin E may be due to its antioxidant effect.

References


1944 Effect of Acrylamide on the Tongue Musculature of Adult Male Albino Rat


تأثير عقار الأكريلاميد على عضلية لسان ذكر الفأر الأبيض البالغ
والدور الوقائي المحتمل لفيتامين E:
دراسة بالمجهرين الضوئي والإلكتروني

المقدمة: تسبب مادة الأكريلاميد مشكلة صحية وبيئية عامة، ينتج التعرض المباشر لمادة الأكريلاميد عن طريق تناول الأطعمة بالكربيهيدرات التي يتم إعدادها في درجات حرارة عالية.

الهدف: هدف هذه الدراسة هو إظهار التأثيرات المجهريه باستخدام المجهرين الضوئي والإلكتروني لمادة الأكريلاميد على عضلة لسان ذكر الفأر الأبيض البالغعدد من مختلفين. مدة قصيرة (عشرين يوما)، وعمر متوسطة (أربعين يوما).

 mentions 10 من ذكور الفأر البالغ في مجموعات. المجموعة الأولى (المضافة مطلق) لم تلق أي دواء. المجموعة الثانية (المضافة ضعيفة) لحقت محلول ملحي طبيعي وفيتامين E في شكل كبسولات. تحتوي كل كبسولة فيه 0.2 مجم من كل المواد، وثانيتها 0.1 مجم في كل كبسولة من كل المواد.

تم إعداد كل كبسولة من الفيتيديوم في جريعة زيتية واحدة، وعشرين يوما. توقفت المجموعة الثالثة فعالية الأكريلاميد. وعشرين يوما. كومب في جريعة زيتية واحدة، وعشرين يوما. توقفت المجموعة السادسة مادة الأكريلاميد، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحدة، عدة مرات يوميا. ييقن الفيتيديوم في جريعة زيتية واحد

نتيجة: بالنسبة لدراسة المجهري الضوئي لعينات من معزول عضلة لسان الفأر البالغ في المجموعة الثالثة فقد كشفت نتيجة تغيرات غير طبيعية. فتهتل نسيج العضلة مع تحليل لخلايا التفاعل التحليلي والخلايا الدموية. و komplexe إعداد إلى ظهور إكمال بسيطة الخلايا وظهور باليهة خلايا. أظهرت معاط السنا المصورة للخلايا الأكبر ريسا في تربة الكريهيدرات حول الأطعمة المضافة وكذلك أظهرت معاط السنا المصورة للخلايا الأكبر ريسا في تربة الكريهيدرات حول الأطعمة المضافة. SLS و PEG بالإنابة لدعم الحالة في حالة الميكروبيواريا، وتشكيل ميكروبيواريا معينة وفرعية يتغير نسيجية هذه التغيرات. إذا تبات الفيتيديوم في المجموعة الرابعة. أدى تبات الفيتيديوم في المجموعة الرابعة إلى التحكم في جميع التغيرات النسيجية المذكورة أعلاه كلاً.

المجموعة الخامسة والأخيرة: يمكن الاستنتاج أن التعرض للأكريلاميد يؤدي إلى تلف عضلة لسان الجرب و الذي يصبح أسوأ مع طول فترة التعرض. يمكن تقليل من هذا التأثير السام من الأكريلاميد عند إعطاء فيتامين E بشكل متلازم معه.